NOIDA INSTITUTE OF ENGINEERING AND TECHNOLOGY, GREATER NOIDA

(An Autonomous Institute)



Affiliated to

DR. A.P.J. ABDUL KALAM TECHNICAL UNIVERSITY, LUCKNOW



Evaluation Scheme & Syllabus

For

M. Tech in Biotechnology (BT) First Year

(Effective from the Session: 2020-21)

NOIDA INSTITUTE OF ENGINEERING & TECHNOLOGY, GREATER NOIDA (An Autonomous Institute)

M.TECH. BIO-TECHNOLOGY

Evaluation Scheme SEMESTER-I

SI.	Subject	Subject	Pei	riods		Eval	uation	Scheme	25	Ene Seme	-	Tota I	Credit
No.	Codes	•	L	т	Ρ	СТ	ТА	TOT AL	PS	TE	PE		
1	AMTBT0101	Applied Biochemistry & Molecular Biology	3	0	0	20	10	30		70		100	3
2	AMTBT0102	Bioprocess Engineering & Technology	3	0	0	20	10	30		70		100	3
3	AMTCC0101	Research Process and Methodology	3	0	0	20	10	30		70		100	3
5		Elective -I*	3	0	0	20	10	30		70		100	3
6		Elective -II*	3	0	0	20	10	30		70		100	3
7	AMTBT0151	Applied Biochemistry & Molecular Biology Lab	0	0	4				20		30	50	2
8	AMTBT0152	Bioprocess Engineering & Technology Lab	0	0	4				20		30	50	2
		TOTAL										600	19

L: Lecture T: Tutorial P: Practical/Project CT: Class Test TA: Teacher's Assessment, ESE: End Semester Examination

(*) Refer the Electives list

Elective-I*

1.AMTBT0111 Immunology & Vaccine Technology

2.AMTBT0112 Quality Assurance and Quality Control

3.AMTBT0113 Applied Clinical Research

Elective-II*

1.AMTBT0114 Biological Treatment of Wastewater

2.AMTBT0115 Nano Biotechnology & Toxicology

3.AMTBT0116 Industrial Biotechnological Products

NOIDA INSTITUTE OF ENGINEERING & TECHNOLOGY, GREATER NOIDA (An Autonomous Institute)

M.TECH. BIO-TECHNOLOGY

Evaluation Scheme SEMESTER-II

SI. No	Subject Codes	Subject	Per	Periods Evaluation Schemes End Semester				Tot al	Cre dit				
			L	Т	Р	СТ	TA	TO TAL	PS	TE	PE		
1	AMTBT0201	Bioinformatics	3	0	0	20	10	30		70		100	3
2	AMTBT0202	Entrepreneurship, IPR &Biosafety	3	0	0	20	10	30		70		100	3
3		Elective – III*	3	0	0	20	10	30		70		100	3
4		Elective- IV*	3	0	0	20	10	30		70		100	3
5		Elective- V*	3	0	0	20	10	30		70		100	3
6	AMTBT0251	Bioinformatics Lab	0	0	4				20		30	50	2
7	AMTBT0252	Entrepreneurship, IPR & Biosafety Lab	0	0	4				20		30	50	2
8	AMTBT0253	Seminar-I	0	0	2				20		30	50	1
		TOTAL										650	20

L: Lecture T: Tutorial P: Practical/Project CT: Class Test TA: Teacher's Assessment, ESE: End Semester Examination

(*) Refer the Electives list

<u>NOIDA INSTITUTE OF ENGINEERING & TECHNOLOGY, GREATER NOIDA</u> (An Autonomous Institute)

Elective-III*

1.AMTBT0211 Genetic Engineering

2.AMTBT0212 Applied Food Biotechnology

3.AMTBT0213 Molecular Modelling & Industrial Application

Elective-IV*

1.AMTBT0214 Bioreactor Analysis & Design

2.AMTBT0215 Enzyme Technology & Industrial Application

3.AMTBT0216 Applied Bioenergy

Elective-V*

1.AMTBT0217 Cell & Tissue Culture Techniques

2.AMTBT0218 Diagnostic Techniques in Biotechnology

3. AMTBT0219 3-D Printing Technology

		M. TECH FIRST YEAR		
Course	Code	AMTBT0101	L T P	Credit
Course	Title	Applied Biochemistry & Molecular Biology	3 0 0	3
Course	objective:			
1	To unde	erstand the various concepts of molecular biology and biochemistry		
2		ne the structure and function of biomolecules and evaluate the comp biomolecules.	olexity of	
3		and the principles of bioenergetics to learn the various pathways.		
4		e the concept of metabolisms of various types.		
5		e structure of genetic material and the central dogma of molecular bio	ology.	
Pre-requ			510851	
110-104	1151105.	Students are expected to have knowledge of basic biology, cell biol- biochemistry	ogy and	
Course	Contents	/ Syllabus:		
Unit 1	Struc	tures and functions of Bio-molecules:		8 hr
	acids, function	hydrates: classification, mono, di, oligo and polysaccharides. Lipic simple, complex & derived lipids. Protein: Amino Acids Struct on, Protein Structure Hierarchy. Nucleic acids: nucleosides, nucleotic & RNA.	ure and	
Unit 2	Bioen	ergetics:		8 hr
	Energ	iew of principles of bioenergetics (free energy, enthalpy and ex y relationships between catabolic and anabolic pathways. Phosphory ers and ATP, Free-energy change for ATP hydrolysis.		
Unit 3	Metal	bolism:		8 hr
	Cycle	lysis, Gluconeogenesis, Respiration and Introduction to the Citric Ac, Electron Transport, Oxidative phosphorylation, Fatty Acid Catacid oxidation, Protein Metabolism: The Urea Cycle		
Unit 4	Gene	Structure and Function		10 hr
	DNA replica	structure, DNA & RNA as a genetic material, RNA World, packa as chromosome, DNA replication- Prokaryotic and eukaryotic ation, Mechanism of replication. Telomeres, telomerase and end replic of telomerase in aging and cancer.	c DNA	
Unit 5	Centr	al Dogma		10 hr
		cription, genetic code, reverse transcription, mRNA processing. Tra regulation, operons: Lac operon, Trp operon, transposons.	anslation,	
Course	outcome			
CO1		After completion of the course, students will understand about the and function of biomolecules	structure	
CO2		They will learn about principles of bioenergetics.		
CO3		They will understand the different types of metabolisms.		
CO4		Students will learn the overall gene structure and function.		
CO5		Students will be able to understand the molecular functioning of cel	lls.	
Text boo	oks			
1		Biochemistry- L.Stryer, Third Edition		
2		Biochemistry- Voet&Voet.		

3	Principles of Biochemistry- A.Lehninger, CBS Publishers and Distributors,				
	1987.				
Reference Books					
1	Watson. J. D, Baker. T. A, Bell. S. P, Gann. A, Levine. M, Losick. R. Molecular Biology of Gene. 6th The Benjamin / Cummings Pub. Co. Inc, 2008.				
2	Darnell, Lodish and Baltimore. Molecular Cell Biology, Scientific American Publishing Inc, 2000.				
3	Bruce Alberts, Alexander Johnson, Julian Lewis, Martin Raff, Keith Roberts, Peter Walter. Molecular biology of the Cell. 4th ed. Garland publishing Inc, 2002				
Journal/Research Paper Link:					
	As suggested by concern subject faculty				

Course (Code AMTBT0102 L T P	Credit
Course 7		3
Course ob	jective:	
1	To provide basic concepts of bioprocess engineering.	
2	To learn engineering principles that can be applied to processes involving cell or enzyme.	
3	To learn the basics of bioreactor design and operation control.	
4	To analyze variety of bioprocess techniques and also conduct related experiments.	
5	To understand various unit operations in bioprocess.	
Pre-requis	sites:	
	Students are expected to have knowledge of basic biology, cell biology and biochemistry	
	ontents / Syllabus :	
UNIT I	Introduction to Bioprocess Technology	8 Hr.
	fermentation processes, Basic design and construction of fermenter and ancillaries, Main parameters for monitoring & control of fermentation processes, Different raw materials used in fermentation industry and their pretreatment, Medium for plant cell culture and animal cell culture, Medium design of commercial media for industrial fermentations-Plackettburman design, response surface methodology, simplex design.	
UNIT II	Stoichiometry of Cell growth	8 Hr.
	Stoichiometry of Cell growth and product formation, elemental balances, degrees of reduction of substrate and biomass, available electron balances, yield coefficients of biomass and product formation, maintenance coefficients Energetic analysis of microbial growth and product formation, oxygen consumption and heat evolution in aerobic cultures, thermodynamic efficiency of growth.	
UNIT III	Mass Transfer in Bioreactors	8 Hr.
	Mass transfer includes transport phenomena in bioprocesses, Factors affecting oxygen transfer rate in bioreactors, Techniques for measurement of volumetric oxygen transfer coefficient, Fluid rheology and factors affecting bioreactor processes, Flow Patterns in agitated tanks, Mechanism & Power requirements of mixing, Scale up of mixing systems.	
	Metabolic Regulation	10 Hr.
UNIT IV		
UNIT IV	Different regulatory mechanisms involved in controlling the catabolic and anabolic processes of microbes, Induction, nutritional repression, carbon catabolite repression, Crabtree effect, feedback inhibition and feedback repression, Concept of Overproduction of metabolites, Case studies on production of Lactic acid, Glutamic acid, Penicillin, Microbial Lipase and Protease, Recombinant Insulin, Interferons, Hepatitis Vaccines etc. Case studies should deal with strain improvement, medium designs, process optimization technology.	

	Unit Operation: Filtration, filter aids, filtration Equipment and filtration theory, Centrifugation process and its equipments, Cell disruption, Aqueous Two-Phase Liquid Extraction. Adsorption process and its operations, Chromatography: Theory and mechanism, Scaling-up chromatography.
Course o	utcome:
CO1	Describe the underlying principles of main bioprocess unit operations like fermentation, downstream processing.
CO2	Demonstrate Stoichiometry of Cell growth and product formation.
CO3	Design or Select appropriate bioreactor models based upon bioproducts and cell lines and other process criteria.
CO4	Develop a basic understanding of regulatory mechanisms involved in controlling the catabolic and anabolic processes of microbes.
CO5	Acquire a basic understanding of various unit operations in bioprocess engineering.
Text bool	3
1	Principles of fermentation technology" by P F Stanbury and A Whitaker, Pergamon press.
2	Bioprocess Technology - Kinetics & Reactors" by A Moser, Springer- Verlag.
3	Biochemical Engineering and Biotechnology Handbook" by B. Atkinson & F. Mavituna, 2nd Ed. Stockton Press.
4	Bioprocess Engineering Principles" by Pauline M. Doran, Academic Press.
5	Biochemical Engineering- S. Aiba, A.E. Humphray, University of Tokyo Press.
Reference	
1	Lee J.M, Biochemical Engineering 2nd ed, Prentice Hall, 2000.
2	Principles of Cell Energetics": BIOTOL series, Butterworth - Heinemann.
3	Biotechnology" Vol.4 Meaning Modelling and Control Ed. K.Schugerl, VCH (1991).
4	Unit operations of Chemical Engineering" 5th ed. by W L McCabe, J C Smith and P. Harriot Mc Graw-Hill (1993).
5	Diffusion" by E L Cussler, Cambridge University Press (1984).
6	Bioprocess Engineering Principles" by Pauline M.Doran, Academic Press.
Journal/I	Research Paper Link:
	As suggested by concern subject faculty

	M. TECH FIRST YEAR	
Course Code	AMTCC0101 L T	P Credit
Course Title	Research Process & Methodology3 0	0 3
Course Object	tive:	
1	To explain the concept / fundamentals of research and their types	
2	To study the methods of research design and steps of resear process	ch
3	To explain the methods of data collection and procedure of sampli techniques	ng
4	To analyze the data, apply the statistical techniques and understa the concept of hypothesis testing	nd
5	To study the types of research report and technical writing.	
Pre-requisites	: Basics of Statistics	
	Course Contents / Syllabus	
UNIT-I	INTRODUCTION TO RESEARCH	8 hours
	tive and motivation of research, types and approaches of research,	
	lied vs. Fundamental, Quantitative vs. Qualitative, Conceptual	
	s versus Methodology, significance of research, criteria of good resea	arch.
UNIT-II	RESEARCH FORMULATION AND DESIGN	8 hours
objective of Liter and identifying t	and steps involved, Definition and necessity of research problem. If rature review, Locating relevant literature, Reliability of a source, W the research problem, Literature Survey, Research Design, Metho	riting a survey
design.		1
UNIT-III	DATA COLLECTION	8 hours
primary and seco	Data, accepts of method validation, Methods of Data Collection ondary data, sampling, need of sampling, sampling theory and Tech different types of sample designs, ethical considerations in research.	
UNIT-IV	DATA ANALYSIS	8 hours
appropriate statis statistical inferen	ations, Data analysis, Types of analysis, Statistical techniques an stical technique, Hypothesis Testing, Data processing software (ence, Chi-Square Test, Analysis of variance(ANOVA) and co fonitoring Research Experiments, hands-on with LaTeX. TECHNICAL WRITING AND REPORTING OF RESEARCH	.g. SPSS etc.) variance, Data
V 1	rch report: Dissertation and Thesis, research paper, review	,
Indexing, cita SCI/SCIE/ESCI/S their ranking, pla right, royalty, tra	conference presentation etc., Referencing and referencing styles, Rest tion of Journals and Impact factor, Types of SCOPUS/DBLP/Google Scholar/UGC-CARE etc. Significance of c giarism, IPR- intellectual property rights and patent law, commerc ade related aspects of intellectual property rights (TRIPS); schola and design of research paper, reproducibility and accountability.	of Indexing- onferences and ialization, copy
	ne: Upon completion of the course, the student will be able to	
CO 1	Explain concept / fundamentals for different types of research	K1
CO 2	Apply relevant research Design technique	К3
CO 3	Use appropriate Data Collection technique	K3
CO 4	Evaluate statistical analysis which includes various parametric to and non-parametric test and ANOVA technique	est K5

CO 5	Prepare research report and Publish ethically.	K6
Text books	· · · · · · · · · · · · · · · · · · ·	
	thari, Gaurav Garg, Research Methodology Methods and Techniques nal publishers, Third Edition.	, New Age
2. Ranjit Ku SAGE 20	mar, Research Methodology: A Step-by-Step Guide for Beginners, 2005.	2 nd Edition,
3. Deepak Cl	nawla, NeenaSondhi, Research Methodology, Vikas Publication	
Reference Boo	oks	
1. Donald Co	oper & Pamela Schindler, Business Research Methods, TMGH, 9 th edition	on
2. Creswell, J sage publications	ohn W.,Research design: Qualitative, quantitative, and mixed methods a ,2013	pproaches

	M. TECH FIRST YEAR	
Course Code	AMTBT0151 L T P	Credit
Course Title	Applied Biochemistry & Molecular Biology Lab 0 0 4	2
Course objectiv	e:	
1	To understand the various concepts of molecular biology and biochemistry	
2	Determine the structure and function of biomolecules and evaluate the complexity of various biomolecules.	
3	Understand the principles of bioenergetics to learn the various pathways.	
4	Evaluate the concept of metabolisms of various types.	
5	Evaluate structure of genetic material and the central dogma of molecular biology.	
Pre-requisites:		
	Students are expected to have knowledge of basic biology, cell biology and biochemistry	
Suggested list of	f Experiment :	
Sr. No.	Name of Experiment	CO
1	Quantitative estimation of amino acids by ninhydrin reaction.	12
2	Quantitative estimation of proteins.	12
3	To separate lipids with the help of thin layer chromatography (TLC).	12
4	To verify the Lambert Beer's law with the help of UV absorption spectra of proteins.	12
5	Protein purification by ammonium sulfate precipitation.	12
6	Isolation of DNA and RNA from animal tissue and plant tissue.	12
7	Gel electrophoretic analysis of various DNA and their restriction digests	1
8	Transformation with plasmid and bacteriophage DNA	13
9	Restriction mapping of plasmid DNA	3
10	Blotting: northern blotting, southern blotting	3
11	PCR technique	3
Lab Course Ou	itcome:	
CO 1	Students will be able to understand the various biomolecules.	
CO 2	Students will learn through demonstration the process of isolation and analysis of different biomolecules.	
CO 3	They will learn about the structure and function of DNA, RNA and Protein.	
CO 4	Students will learn advanced molecular methods.	

	M. TECH FIRST YEAR	
Course Code	AMTBT0152 L T P	Credit
Course Title	Bioprocess Engineering & Technology Lab0 0 4	2
Course objectiv	a•	
1	To understand the various concepts of microbial culturing.	
2	To learn the activation energy, volumetric oxygen transfer coefficient etc.l	
3	To Understand the principles and various pathways of enzyme production.	
4	Evaluate the concept of separation and purification of microbial produce.	
5	To understand the process of fermentation.	
Pre-requisites:	~	
	Students are expected to have knowledge of basic biology, cell biology and biochemistry	
Suggested list of	f Experiment :	
Sr. No.	Name of Experiment	
1	Determination of kinetic parameters for batch cultivation of yeast under shake flask conditions.	
2	Determination of volumetric oxygen transfer coefficient (KLa)	
3	Determination of activation energy (Ea) of microbial strains.	
4	Process optimization for enzyme production using specific experimental design.	
5	Preparation of immobilized enzymes & cells and evaluation of kinetic parameters.	
6	Computational Design of Fermentative Process.	
7	Fermenter designing and the study of various parts of fermenter and their function for microbial cell culture.	
8	Fermentative production of Penicillin by using <i>Peniciliumchrysogenum</i> .	
9	Microbial production of enzymes Cellulase & Protease.	
10	Ethanol production from molasses or starchy raw material.	
11	Fermentative production of Wine from grapes.	
12	Separation and purification of microorganisms from yogurt and cheese.	
13	Fermentative production of alpha amylase under solid & submerged conditions	
14	Protein profiling of fermentation broth through dialysis procedure.	
15	To study the Scale-up and Sterilization in Bioreactors	
Lab Course Ou	tcome:	
CO 1	Student will be able to understand the various concepts of microbial culturing.	
CO 2	Student will learn the activation energy, volumetric oxygen transfer coefficient etc.	
CO 3	Student will Understand the principles and various pathways of enzyme production.	
CO 4	Student will be able to evaluate the concept of separation and purification of microbial produce.	
CO 5	Student will be able to understand the process of fermentation.	

	M. TECH FIRST YEAR	
Course Code	AMTBT0111 L T P	Credit
Course Title	Immunology & Vaccine Technology3 0 0	3
Course objective	<u> </u>	
1	Learn the concept and components of the Immune system.	
2	Understand the kinetics and mechanisms of immune response.	
3	Evaluate the concept of vaccination and various types of vaccines.	
4	Understand the concept of various vaccine types viz. viral vaccines, bacterial vaccines and parasitic vaccines etc.	
5	Understand the vaccine industry and the safety and legal issues related to its production.	
Pre-requisites:		
Course Contonto	Students are expected to have knowledge of basic Cell and Molecular biology, knowledge of the various diseases and causative agents will be an edge.	
Course Contents		0.1
UNIT-I	Fundamental of Immune System	8 hr
	Fundamental concepts and anatomy of the immune system, Components of innate and acquired immunity, Humoral and Cell mediated immunity, Hematopoiesis, Antigens, immunogens, haptens, Major Histocompatibility Complex - MHC genes, MHC and immune responsiveness and disease susceptibility, HLA typing.	
UNIT-II	Immunological Processes	8 hr
	Immunoglobulins-basic structure, classes and subclasses of immunoglobulins, antigenic determinants, Multigene organization of immunoglobulin genes, Immunological basis of self –non-self discrimination; Kinetics of immune response, memory; B cell maturation, activation and differentiation; Generation of antibody diversity, Antigen processing and presentationendogenous antigens and exogenous antigens.	
UNIT-III	Basic Introduction to Vaccines	8 hr
	A short history of vaccination, Active and passive immunization, General immunization practices, Vaccination of immunocompromised hosts, Vaccination of human immunodeficiency virus infected persons, Vaccines, Live, killed, attenuated, subunit vaccines; Vaccine technology- Roleand properties of adjuvants, recombinant DNA and protein-based vaccines, plant-based vaccines, reverse vaccinology; Peptide vaccines, conjugate vaccines.	
UNIT-IV	Recent Advances in Vaccines	10 hr
	Licensed vaccines, Viral Vaccine (Poliovirus vaccine-inactivated & Live, Rabies vaccines Hepatitis A & B vaccines), Bacterial Vaccine (Anthrax vaccines, Cholera vaccines, Diphtheria toxoid), Parasitic vaccine (Malaria Vaccine).	
UNIT-V	Vaccine Industry (Production & Regulations)	10 hr
	The vaccine industry, Vaccine manufacturing, Evolution of adjuvants across the centuries, Vaccine additives and manufacturing residuals, Regulation and testing of vaccines, Regulation of vaccines in developing countries, Vaccine safety and Legal issues.	
Course outcome	-,	

CO 1	After completion of the course, students will understand the fundamentals of the immune system.	
CO 2	They will learn about immunological processes.	
CO 3	They will understand the different types of immunization and vaccines.	
CO 4	Students will learn the different types of advanced vaccines.	
CO 5	Students will be able to understand the vaccine industry and their production process.	
Text books		
1	Kuby, RA Goldsby, Thomas J. Kindt, Barbara, A. Osborne Immunology, 6th Edition, Freeman, 2002.	
2	Brostoff J, Seaddin JK, Male D, Roitt IM., Clinical Immunology, 6th Edition, Gower Medical Publishing, 2002.	
3	Janeway et al., Immunobiology, 4th Edition, Current Biology publications., 1999. 4. Paul,	
4	Fundamental of Immunology, 4th edition, Lippencott Raven, 1999.	
Reference Books	S	
1	Stanley A. Plotkin & Walter Orenstein & Paul A. Offit, Vaccines, 6th Edition 2013 BMA Medical Book Awards Highly Commended in Public Health! Elsevier Publication.	
2	Roitt's Essential Immunology. 11th ed. P. Delves, et al., ed., Blackwell Publishing, 2006.	
3	Bruce Alberts, Alexander Johnson, Julian Lewis, Martin Raff, Keith Roberts, Peter Walter. Molecular biology of the Cell. 4th ed. Garland publishing Inc, 2002	
Journal/Researc	ch Paper Link:	
	As suggested by concern subject faculty	
L		

~		M. TECH FIRST YEAR		<u> </u>
	se Code		LTP	Credit
	se Title	Quality Assurance and Quality Control	3 0 0	3
Cours	se objectiv			
1		the basics of GLP		
2		the manufacturing process and its audit.		
3	To under	rstand the clinical trial process		
4	11.5	the statistical tools to the various QC events		
5	To under	rstand the tools and softwares used in QC and QA.		
Pre-re	equisites:			
Stude	ents a	re expected to have knowledge of basic biology, cell biology	and biochemistry	
Cours	se Conten	ts / Syllabus :		
TINIT				
UNIT		ncept of Quality control and quality assurance	Total Orgalita-	
		ncept and evolution of quality control and quality assurance. Tanagement, Philosophy of GMP and CGMP. Quality control la	~ •	
		ponsibilities: GLP protocols on nonclinical testing control on		
	dat	a generation, integration and storage, standard test procedure,		
	Ũ	idelines.		
UNIT		cumentation practices and root cause analysis		
	fini act	tention of sample records, Quality review and batch release do ished products, Good documentation practices, route cause an ion preventive action (CAPA), out of specifications (OOS) an OT)	alysis, corrective	
UNIT	III Co	ncept of Audits		
	auc qua	nual product quality review and parametric release, Audits,Pr dit, conducting audit, Audit Analysis, Audit Report and Audit ality audits of manufacturing processes and facilities, audits of se Studies of Audit reports.	follow up,	
UNIT	'IV Qu	ality agreements and risk management		
	qua	ncepts and management of contract manufacturing guidelines ality risk management, ICH guidance for industry, BABE (bio bequivalence) studies, post marketing surveillance, Pharmacov	availability and	
UNIT	V To	ols and softwares in QC and QA		
	Co app sol	ttistical Tools for Quality Control and Precision, Tools of Pro- ntinuous Improvement. Softwares for inspection and quality t plications. concept of automation of procedure through Digita utions. Systematic approach to scale-up and technology trans otechnology quality systems: Applications and challenges.	esting and their al, IoT and BOTS	
Cours	se outcom	e		
CO1		cognize the importance of quality control and assurance and uncept of GMP, CGMP and GLP.	inderstand the	
CO2		cognize the importance of good documentation practices and a eventive actions.	reframe the	
CO3		alyse, develop, follow and audit the quality standards and guid lowed in a biotechnology industry.	delines, being	

CO4	Understand the contract guidelines to effectively manage the quality agreements.	
CO5	Apply statistical tools and modern software to evaluate and ensure quality control, assurance and precision.	
Text books		
1	Sharp J. Good Pharmaceutical Manufacturing Practice: Rationale and Compliance. CRC Press; 2005.	
2	Gad SC. Pharmaceutical Manufacturing Handbook: Production and Processes. John Wiley & Sons; 2008.	
3	Steinborn L. GMP/ISO Quality Audit Manual for Healthcare Manufacturers and Their Suppliers, Sixth Edition, (Volume 1 - With Checklists and Software Package). Taylor & Francis; 2003.	
4	Kolman J, Meng P, Scott G. Good Clinical Practice: Standard Operating Procedures for Clinical Researchers. Wiley; 1998.	
5	Waller P. An Introduction to Pharmacovigilance. John Wiley & Sons; 2011.	
Reference l	Books	
1	Niazi S. Handbook of Bioequivalence Testing. CRC Press; 2007.	
2	Chalmers AA. International Pharmaceutical Registration. Interpharm Press; 2000.	
3	Edwards AJ. ISO 14001 Environmental Certification Step- by-Steps: Revised Edition. Butterworth-Heinemann; 2003.	
4	Mantus D. FDA Regulatory Affairs: A Guide for Prescription Drugs, Medical Devices, and Biologics. Informa Healthcare USA; 2008.	
5	Chalmers AA. International Pharmaceutical Registration. Interpharm Press; 2000.	
6	Contract manufacturing arrangement for drugs, quality agreements: guidance for industry,November 2016.	
Journal/Re	search Paper Link:	
	As suggested by concern subject faculty	

	M. TECH FIRST YEAR		
Course Code	AMTBT0113 L	T P	Credit
Course Title	Applied Clinical Research3	00	3
Course objectiv	/o•		
1	To learn the basic of drug development process		
2	To learn the basic step involve in clinical trial of drug.		
3	To understand the ethics involved in clinical research		
3 4	To understand the principles of controlled clinical trials		
5	To apply the statistical tool for data management.		
<u></u>	To upply the statistical tool for data management.		
	Students are expected to have knowledge of basic biology, cell biology a biochemistry	ind	
Course Content	ts / Syllabus :		
UNIT I:	Introduction to clinical research		8 Hr.
	Basic pharmacology and drug development process, cl researchdefinition, Basic terminology used in clinical research, precl studies, Introduction to pharmacoeconomics, Types of clinical trials, s blinding, double blinding, open access, randomized trials and their exami interventional study, Good Clinical Practices, Types and Scope of Cl Research.	single nples,	
UNIT II:	Clinical trials		8 Hr.
	New drug discovery process- purpose, main steps involved in new discovery process, timelines of each steps, advantages and purposes of steps, Pre clinical toxicology: General principles, Systemic toxic (Single dose and repeat dose toxicity studies), Carcinogenicity, Mutager Teratogenicity, Reproductive toxicity, Local toxicity, Genotoxicity, a toxicity requirements, Phase-I, II, III, IV trials: Introduction and desig Various phases of clinical trials, Post Marketing surveillance, method allocation and randomization, Informed consent process in brief monit treatment outcome, Termination of trial, Safety monitoring in clinical trials	f each ology nicity, nimal gning, ods & ls of oring,	
UNIT III:	Ethics & Regulations in Clinical research		8 Hr.
	Ethical Theories and Foundations, Ethics Review Committee and Info Consent Process, Integrity& Misconduct in Clinical Research, unethical thalidomide tragedy, Conflicts of Interest, Evolution and Histon Regulations in Clinical Research, Study of various clinical trials (comp or ongoing), Patents US Regulatory Structure, Clinical Trial Application India Import & Export of Drug in India, Investigational New Drug application (IND), New Application (NDA), Abbreviated New Drug Application (ANDA), Post Approval Activities, PMS, FDA Audits and Inspections EU Regu Affairs, EMEA Organization and Function, INDIAN Regulatory sy	trials, y of oleted on in Drug Drug latory	
	Schedule Y- Rules and Regulations.		

	Clinical trial design (observational and interventional) protocol, consent in clinical trials, placebo, bias and methods to prevent bias, monitoring. Multicentre clinical trials, Requirements, regulations and feasibility, Designing of Protocol, CRF, eCRF, IB, ICF, SOP BA/BE Studies Report writing, Publication, Improving patient enrolment and retention in Clinical Trials. ADR monitoring,Pharmacovigilance Training in clinical research.	
UNIT V	: Biostatistics and data management	10 Hr.
	 Preparation of a successful clinical study, Study management, Project management Documentation, Monitoring, Audits and Inspections. Budgeting in clinical research, Supplies and vendor management. Importance of statistics in clinical research Statistical considerations at the design, analysis and reporting stage. Data management, Data validation, SAE reconciliation, query management Software considerations. Clinical Trial studies: Cancers and Other Neoplasms, Behaviours and mental Disorders, Immune System studies, Urinary Tract, Sexual Organs and pregnancy condition. 	
Course	outcome	
CO1	Describe the process of drug development and principles of clinical pharmacology.	
CO2	Develop a clear understanding of why ethics are important in clinical research and be familiar with the regulatory practices in place to protect both the researcher and the subject	
CO3	Effectively manage the regulatory process from Innovation \rightarrow Discovery \rightarrow Approval \rightarrow Commercialization to bring the product to the market globally.	
CO4	Communicate ideas and data in writing, including of scientific concepts and research design of clinical trials	
CO5	Describe the various types of clinical studies and the methods used to choose the appropriate design, evaluation and interpretation of clinical trial results.	
Text bo	bks	
1	Basic and Clinical Pharmacology, Prentice hall, International, Katzung, B.G.	
2	Clinical Pharmacology, Scientific book agency, Laurence, DR and Bennet PN.	
3	Clinical pharmacokinetics, Pub. Springer Verlab, Dr. D.R Krishna, V. Klotz	
4	Remington Pharmaceutical Sciences, Lippincott, Williams and Wilkins	
5	Drug interaction, Kven Stockley. Hamsten	
	ce Books	
1	Clinical pharmacology and drug therapy Grahame smith and Aronson,	
2	Text Book of Therapeutics Drug and Disease Management Hardbound. Richard A Helms,	
3	Clinical Pharmacy and therapeutics Herfindal E T and Hirschman JL, Williams and Wilkins,	
4	Methodology of Clinical Drug Trials, 2nd Edition. Spriet A., Dupin-Spriet T., Simon P. Publisher: Karger.	
Journal	Research Paper Link:	
	As suggested by concern subject faculty	

~	~ .	M. TECH FIRST YEAR	LTD	<u> </u>
Course		AMTBT0114	LTP	Credi
Course	Title	Biological Treatment of Waste Water	300	3
Course o	biective:			
1		about the mass balance involved in waste water treatment		
2	To under	stand the anaerobic treatment process.		
3		about the various chemical and physical processes involved in wa	ste water	
4	To under	stand the basic of phosphorus and nitrogen removal		
5	To Learn	about the recycling of waste		
Pre-requ	Students biochemi	•	1	
Course C	Contents /	Syllabus:		
UNIT I-	SEL	IVATED SLUDGE PROCESS-PROCESS ANALYS ECTION		8 Hr.
	Mass Anae (TSU	acteristics of Activated Sludge (aerobic and anaerobic); Analysis of Balance Analysis. Reactors used in waste water treatment- Up Fl robic Sludge Blanket (UASB), Two-stage, Aerobic UNI Tank Sys System, Route Zone Treatment, Submerged Aerobic Fixed Film (tor, and Fluidized Aerobic Bioreactor (FAB).	ow stem	
UNIT II-	· AER	OBIC FIXED-FILM & ANAEROBIC TREATMENT PROC	ESSES	8 Hr.
	Rotat Circu Anae	Im process considerations; Trickling Filters and Biological Tower ing Biological Contactors; Granular – Media Filters; Fluidized – Ilating Bed- Biofilm reactors. Hybrid Biofilm/suspended growth p robic Processes: Methanogenesis, process chemistry and microbic ess kinetics and factors for the design of anaerobic digestors.	Bed & rocesses.	
UNIT III	- ADV	ANCED WASTE WATER TREATMENT		8 Hr.
	Remo Filtra proce	nologies used in advanced treatment-Classification of technologie oval of Colloids and suspended particles-Depth Filtration, Surface tion, Membrane Filtration Absorption, Ion Exchange, Advanced o ess, Activated Carbon, Air Stripping, Heavy Metals Removal, Stea- ping, Chemical Precipitation, and Electrolysis.	oxidation	
UNIT IV		LOGICAL PHOSPHORUS AND NITROGEN REMOVAL		10 Hr.
	Nitrif sludg Denit Biom	fication & Denitrification Processes: Biochemistry and Physiology ying Bacteria; Common process considerations; One sludge versu e nitrification. Physiology of Denitrifying Bacteria; Tertiary trification; One- sludge denitrification, Normal Phosphorus Uptak ass; Mechanism for Biological Phosphorus Removal; Enhanced E phorus Removal by Bacteria and Algae.	is two e into	
UNIT V	-	IRONMENTAL CONCERNS & RECYCLING OF WASTES		10 Hr.
	Envir Laws Audit	ronmental regulations and technology- Regulatory Concerns, Tech , regulations and permits, Air, Water, Solid Waste, Environmenta ting, National Environmental Policy act, Occupational Safety and OSHA), Storm Water Regulations; Technology (waste water); Re	nnology; l Health	

CO1	After completing the course students will able to perform mass balance for the bioreactor
CO2	After completing the course students will able to design an anaerobic system
CO3	After completing the course students will able to categorize various chemical and physical processes involved in waste water treatment.
CO4	After completing the course students will able to describe the basic of phosphorus and nitrogen removal
CO5	After completing the course students will able to perform recycling of waste
Text books	
1	Wastewater Engineering: Treatment Disposal Reuse by Metcalf & Eddy
2	Environmental Biotechnology : Principles and Applications by Bruce E. Rittmann
3	Waste water Engineering Treatment and Reuse: McGraw Hill, G.
4	Industrial Waste Water Management Treatment and Disposal by Waste Water McGraw Hill III Edition 2008.
5	Biological Wastewater Treatment", Second Edition, Marcel Dekker, Inc., New York,
Reference B	ooks
1	Introduction to Waste Water Treatment- R. S. Ramalho, Academic Press.
2	Environmental Biotechnology, B.C. Bhattacharya &Ritu Banerjee, Oxford Press, 2007.
Journal/Res	earch Paper Link:
	As suggested by concern subject faculty

		M. TECH FIRST YEAR		
Course (Code	AMTBT0115	L T P	Credit
Course 7	ſitle	Nano Biotechnology & Toxicology	300	3
Course ob	jective:			
<u>1</u>	jeenve.	To understand the fundamentals concepts of nanotechnology		
2		To learn about the different types of nanoparticles		
3		To understand the principle behind the different char techniques involved in nanotechnology	acterization	
4		To understand the applications of nanotechnology		
5		To learn the toxicology of nanomaterials		
Pre-requis	sites:			
		Students are expected to have knowledge of basic biology, cell and biotech	l biology	
Course Co	ontents / S	Syllabus :		
UNIT-I		oduction to Nanobiotechnology:		8hr.
	Botto Curre	nition of Nanobiotechnology, History, Origin, Fundamental om-up versus Top-down approaches, Discussion on Nanof ent research, Tool and Techniques, Applications and Implication of abrication.	abrication,	
UNIT-II	Nano	omaterials and Nanoparticles:		8hr.
	Buck Silve	on nanotubes and related structures, Properties, Synthesis, Apply balls, Nanoparticles types and their synthesis, Application r and Zinc oxide nanoparticles, Interaction of nanopart embrane and genes.	n of Gold,	
UNIT-III	Nano	ocharecterization Tool and Techniques:		8hr.
	Scan (STM	visible spectrophotometry, Fourier transform infrared spectrosconing Electron Microscopy (SEM), Scanning tunnelling r (1), Transmission electron microscopy (TEM), Atomic force r (1), Zeta Potential size analyser etc.	nicroscopy	
UNIT-IV	Nano	omedicine and Sensor Technology:		10 hr.
	Drug	delivery tools, Bioavailability, Nano imaging agents, Protein a ery (Cancer and Surgery) and Nano sensors technology with app		
UNIT-V	Toxi	cology:		10 hr.
	Toxic toxic mech	nition of toxicology, History and origin of toxicology, Pricology, Concept of Toxicology, Types of toxicology, Na ity evaluation mechanism as in vitro, Nanomaterial toxicity nanism as in vivo, Assessment of nanoparticles toxicity: A otoxicity, Genotoxicity, Hepatotoxicity, Neurotoxicity, Nep	evaluation case study	
Course ou	/			
CO1	After co	mpleting this course, the students will be able to learn the fu of nanotechnology	ndamentals	
CO2		npleting this course, the students will be able to ability for under rentiate the various nano materials	rstanding	
CO3		mpleting this course, the students will be able to understand the different characterization techniques involved in nanotechnological structures involved in the students of t	· ·	

CO4	After completing this course, the students will be able to get insight the application of nanotechnology in drug delivery system	
CO5	After completing this course, the students will be able to evaluate the toxicology of nanomaterials	
Text boo	ks	
1	Nanomedicine: Biocompatibility- Robert A. Freitas; Landes Biosciences	
2	The Nanobiotechnology Hand Book- YobingXie, CRC Press.2012	
3	Nanobiotechnology: Christof M. Niemeyer, Chad A. Mirkin, John Wiley & Sons, 2004	
Referenc	e Books	
1	Nancy A. Monteiro-Riviere, C. Lang Tran., 'Nanotoxicology: Characterization, Dosing and Health Effects', Informa Healthcare publishers, 2007.	
2	P. Houdy, M. Lahmani, F. Marano, 'Nanoethics and Nanotoxicology', SpringerVerlag Berlin Heidelberg 2011.	
Journal/l	Research Paper Link:	
	As suggested by concern subject faculty	

		M. TECH FIRST YEAR		
Course	Code	AMTBT0116	L T P	Credit
Course	Title	Industrial Biotechnological Products	3 0 0	3
<u> </u>				
Course o	-			
1		n about the different media for the growth of microbes	halitaa	
2		erstand the production process of Primary and Secondary meta		
3		gn and deliver useful modern biotechnology products to the S tand the methods to obtain enzymes of industrial importance	-	
4		broduct development Research &Development	e and in general	
5	-	erstand the manufacturing of various organic and alcoholic pro-	oducts	
Pre-requ		6 6 1		
1	Stude	ents are expected to have knowledge of basic biology, miggy and biochemistry	crobiology, cell	
Course C	Contents	/ Syllabus :		
Unit I	Fund	amentals of Fermentation		8 hr.
	Indus	rent types of culture media; Substrates for industrial micro trially important micro-organisms: Isolation, screening, Select ess optimization techniques.	.	
Unit II	Prod	uction of Metabolites		8 hr.
	ethan wine	ess technology for the production of various Products: Prima ol, citric acid, vinegar and amino acid; Production of alcoh and beer; Secondary metabolites: Antibiotics; Process tech action of microbial biomass.	olic beverages:	
Unit III	Biop	roducts		8 hr.
	Produ	duction and production of secondary metabolites with sor action of bioplastics (PHB, PHA), bioinsecticides, olymers, Biofertilizers and biological weapons with reference t	bioherbicides,	
Unit-IV	Prod	uction of industrially important enzymes		8 hr.
	ferme Prote	action of industrially important enzymes: Solid state fermentat entation, Extraction, Purification and characterization of indu ases, Cellulase, Lipase, Amylase and Pectinase, industrial nes for production of drugs and fine chemicals, Enzyme based	strial enzymes: process using	
Unit V	Prod	uction of Fermented Food Products		8 hr.
	comr prepa bever	nological processes for industrial manufacture of select mercial importance from plants and animal sources. Proce tration of Yoghurt, acidophilus milk, Koumis, kefir, cheese, b rage, vinegar and oriental fermented food. Food packagin ved in the commercially important food processing methods.	ss involved in pread, alcoholic	
Course o				
CO1		p key practical skills in fermenting biotechnology and be ons and commercial opportunities in fermentation-based biotec		
CO2		e their understanding that 'industrial biotechnology' is be to control the growth of microorganisms	based on using	
CO3	Develo	p knowledge of a variety of fermentation strategies		
CO4	A	e potential business opportunities in fermentation-based biotec	hnology	

CO5	Explore the biological and technological principles which govern actual and potential bio-business	
Text boo	bks	
1	Industrial Microbiology, Casida Jr. L. E. 1968) new Age International (P) Ltd. New Delhi.	
2	Presott& Dunn's Industrial Microbiology. Ed. E.G. Reed (1987). CBS Publishers, New Delhi.	
3	Biotechnology: A Text book of Induxctrial Microbiology 2 nd Edition. Crueger, W. and Cruger, A. (2000) Panima Publishing Corporation, New Delhi.	
Referen	ce Books	
1	Enzmes: Biochemistry, Biotechnology, Clinical Chemistry, Palmer, T. (2000) Horwood Publishing Colphon.	
2	Manual of Industrial Microbiology and Biotechnology 2nd Edition. Ed. Arnold L. Demain and Julian E. Davies (1999) ASM Press Washington D.C.	
3	Microbiology, Pelzar Jr. M.J.: Chan E.C.S. and Krieg, N. R. (1993) Tata Mc Graw Hill, New Delhi.	
Journal	Research Paper Link:	
	As suggested by concern subject faculty	

õ		M. TECH FIRST YEAR		
Course C	Code	AMTBT0201	L T P	Credit
Course T	itle	Bioinformatics	300	3
Course ob	iective	:		
1	<u>.</u>	To learn the various online databases		
2		To learn the online tools for analysing various methods of sequend alignment	ce	
3		To understand the phylogenetic analysis and related conclusions		
4		To understand the concepts of system biology.		
5		To understand the various methods of genome sequencing.		
Pre-requis	sites:			
		Students are expected to have knowledge of basic biology, cell bio biochemistry	ology and	
		/ Syllabus		
UNIT I	Bio	logical Databases		8 Hr.
	Spe data	Bank, DDBJ, EMBL, PIR, Uniprot-KB, SWISS-PROT, ccialized databases: Pubmed, OMIM, Medical databases, KE abases; Genome databases at NCBI, EBI, TIGR, SANGER. Ov er popular tools for various bioinformatics exercises.	GG, EST	
UNIT II	Seq			
		uence Alignment		8 Hr.
	scor sigr Asp Mer		Statistical s, Practical Alignment	8 Hr.
UNIT III	scor sigr Asp Met Lov	uence Alignment oduction to sequence alignment, Optimal Alignment Methods, S res, substitution matrices, PAM, BLOSUM, Gap penalties, nificance of Alignments, Pair wise sequence alignment algorithms beet of Multiple Sequence Alignment, Progressive and Iterative thods, CLUSTALW, Database similarity searching, FASTA,	Statistical s, Practical Alignment	8 Hr. 8 Hr.
UNIT III	scor sigr Asp Met Lov Phy Intr Phy Met Util NT	uence Alignment oduction to sequence alignment, Optimal Alignment Methods, S res, substitution matrices, PAM, BLOSUM, Gap penalties, nificance of Alignments, Pair wise sequence alignment algorithms bect of Multiple Sequence Alignment, Progressive and Iterative thods, CLUSTALW, Database similarity searching, FASTA, wComplexity Regions. PSIBLAST, PHI-BLAST.	Statistical s, Practical Alignment BLAST, c Models, d Building mapping, IT, Vector	
UNIT III UNIT IV	scor sigr Asp Met Lov Phy Intr Phy Met Util NT prop	juence Alignment oduction to sequence alignment, Optimal Alignment Methods, S res, substitution matrices, PAM, BLOSUM, Gap penalties, nificance of Alignments, Pair wise sequence alignment algorithms beet of Multiple Sequence Alignment, Progressive and Iterative thods, CLUSTALW, Database similarity searching, FASTA, wComplexity Regions. PSIBLAST, PHI-BLAST. Mogenetic Analysis and Primer Design roduction to Phylogenetic analysis, Elements of phylogenetic Mogenetic Data Analysis, Phylogenetic Tree -construction steps and thods, Case studies related to phylogenetic analysis, Restriction lities, DNA strider, MacVector and OMIGA, gene construction K I, Web based tools (MAP, REBASE); Primer design – Prim	Statistical s, Practical Alignment BLAST, c Models, d Building mapping, IT, Vector	
	scor sigr Asp Met Lov Phy Intr Phy Met Util NT prop Sys Sys	juence Alignment oduction to sequence alignment, Optimal Alignment Methods, S res, substitution matrices, PAM, BLOSUM, Gap penalties, nificance of Alignments, Pair wise sequence alignment algorithms bect of Multiple Sequence Alignment, Progressive and Iterative thods, CLUSTALW, Database similarity searching, FASTA, wComplexity Regions. PSIBLAST, PHI-BLAST. Mogenetic Analysis and Primer Design oduction to Phylogenetic analysis, Elements of phylogenetic Mogenetic Data Analysis, Phylogenetic Tree -construction steps an thods, Case studies related to phylogenetic analysis, Restriction lities, DNA strider, MacVector and OMIGA, gene construction K I, Web based tools (MAP, REBASE); Primer design – Prim grams and software (PRIME3)	Statistical Alignment BLAST, C Models, d Building mapping, IT, Vector her design formatics, databases, of Proteins	8 Hr.

Course CO1	Genome sequencing technology and analysis methods, Bioinformatics tools and automation in Genome Sequencing, analysis of gene expression data, Utility of EST database in sequencing, Bioinformatics in detection of Polymorphisms, SNPs and their relevance, Bioinformatics tools in microarray data analysis. 	
	Students will be able to use the online tools for analysing various macromolecules	
CO2	of the cells	
CO3	The students shall Identify the role of phylogenetic analysis and related conclusions	
CO4	To learn the use of various tools for molecular analysis	
CO5	To understand the various methods for macromolecular sequencing	
Text bo	oks	
1	Bioinformatics (Sequence and Genome Analysis)- David W. Mount, Cold Spring Harbor Laboratory Press, 2001.	
2	Bioinformatics- Zoe Lacroix, Terence Critchlow, Morgan Kaufmann Publishersm, 2004.	
3	Bioinformatics – From Genomics to Drugs, Violume 1; Basic Technoliges, Thomas Lengauer, Wiley- VCH, 2001.	
4	Bioinformatics (Practical Approach): Sequence, Structure and Databanks – Des Higgins, OXFORD Univ. Press, 2003.	
5	Bioinformatics Computer Skills – Gibas&Jambeck, O' Reilly, 2001, I Ed.	
Referen	ice Books	
1	Bioinformatics Computing- Bryan Berjeron, Prentice-Hall of India, Private Ltd., 2003.	
2	Computational Molecular Biology (An Algorithmic Approach)- Pavel A. Pevzner, PrenticeHall of India, Private Ltd., 2004.	
3	11. Introduction to bioinformatics- T K Attwood, D J Parry-Smith, Pearson Education, 2004.	
4	Sequence Analysis (In A Nutshell)- Scott Market & Darryl Leon, O' Reilly, Ist Edition, 2003.	
5	Scolnick. J.; Drug Discovery and Design, Academic Press, London, 2001.	
6	N. R. Cohen, Editor, Guidebook on Molecular Modeling in Drug Design. Academic Press, San Diego, 1996.	
Journa	/Research Paper Link:	
	As suggested by concern subject faculty	

0	M. TECH FIRST YEAR	TTP	C "
Course (Credi
Course 7	Fitle Entrepreneurship, IPR & Biosafety	3 0 0	3
Course ob	niective		
<u>1</u>	To learn the basics of accounting and finance in business		
2	To learn about the various policies of marketing		
3	To understand the use of IT in business development		
4	To learn about the IPR and its legal provisions.		
5	To learn about the various biosafety in various biological syste	ems.	
Pre-requi	sites:		
	Students are expected to have knowledge of basic biology, cel biochemistry	l biology and	
Course Co	ontents / Syllabus		
Unit I	Accounting and Finance:		8 Hr.
	Approach a bank for a loan; Sources of financial assistance; Mak proposal/Plan for seeking loans from financial institution and Bank bank for capital expenditure and for working; Statutory and legal re starting a company/venture; Budget planning and cash flow manag in accounting practices: concepts of balance sheet, P&L account, an bookkeeping; Estimation of income, expenditure, profit, income tax	cs; Funds from quirements for gement; Basics ad double entry	
Unit II	Marketing:		8 Hr.
	Assessment of market demand for potential product(s) of in conditions, segments; Prediction of market changes; Identify customers including gaps in the market, packaging the product; M branding issues; Developing distribution channels; Pricing/Policie Promotion/Advertising; Services Marketing Negotiations/Str financiers, bankers etc.; With government/law enforcement aut companies/Institutions for technology transfer; Dispute resolution s environment/changes; Crisis/ Avoiding/Managing; Broader thinking	ing needs of arket linkages, s/Competition; rategy: With chorities; With	
Unit III	Information Technology:		8 Hr.
	How to use IT for business administration; Use of IT in impro- performance; Available software for better financial manageme setup, management. Human Resource Development (HRD): Lea Managerial skills; Organization structure, pros & cons of differ Team building, teamwork; Appraisal; Rewards in small scale set up.	nt; E-business idership skills; rent structures;	
Unit IV	IPR:		8 Hr.
	Introduction to Intellectual Property, Types of IP: Patents, Tradema & Related Rights, Industrial Design, Traditional Knowledge, Indications, Protection of New GMOs; International framework for of IP IP as a factor in R&D IPs of relevance to Biotechnology Studies; Introduction to History of GATT, WTO, WIPO and TRI	Geographical the protection and few Case	
	patent application and Process of Technology Transfer	PS, Filing of a	

	An Introduction; Historical Backround; Introduction to Biological Safety Cabinets; Primary Containment for Biohazards; Biosafety Levels; Biosafety Levels of Specific Microorganisms; Recommended Biosafety Levels for Infectious Agents and Infected Animals; Biosafety guidelines - Government of India; Roles of Institutional Biosafety Committee, Risk Analysis; Risk Assessment; Risk management and communication; Overview of National Regulations and relevant International Agreements including Cartagena Protocol.	
Course	outcome	
CO1	The students shall get an adequate knowledge on Accounting and Finance and will be able to do budget planning for any new venture	
CO2	Students will be able to Assessment of market demand for potential product(s) of interest and External environment/changes; Crisis/ Avoiding/Managing Broader vision–Global thinking	
CO3	The students shall Identify the role of Information Technology for business growth	
CO4	To disseminate knowledge on patents, patent regime in India and abroad and registration aspects and to make students aware about current trends in IPR and Govt. supports in promoting IPR	
CO5	The students shall Identify the role of regulatory committees in controlling the risk. Students should get enough information on ethical issues linked to research on animal models, transgenic, clinical trials.	
Text bo	oks	
1	Selected papers from scientific journals.	
2	Nithyananda, K V. (2019). Intellectual Property Rights: Protection and Management. India, IN: Cengage Learning India Private Limited.	
3	Neeraj, P., &Khusdeep, D. (2014). Intellectual Property Rights. India, IN: PHI learning Private Limited.	
4	V Sreekrishna, 2017. Bioethics and Biosafety in Biotechnology by New Age International publishers.	
E Refer	rence resources	
	https://kclau.com/wealth-management/best-budgeting-tools-online- softwares/https://www.ccl.org/articles/leading-effectively-articles/fundamental-4- coreleadership-skills-for-every-career- stage/http://www.yourarticlelibrary.com/organization/8-types-of- organisationalstructures-their-advantages-and-disadvantages/22143 https://opentextbc.ca/organizationalbehavioropenstax/chapter/reward-systems- inorganizations/#ch08rfin-9 https://online.hbs.edu/blog/post/accounting-skills-for- entrepreneurshttps://www.investopedia.com/terms/f/feasibility- study.asphttps://www.extension.iastate.edu/agdm/wholefarm/html/c5- 92.htmlhttps://economictimes.indiatimes.com/wealth/tax/how-to-compute-your- totaltaxabl e-income/articleshow/52956796.cms?from=mdr	
	•Subramanian, N., &Sundararaman, M. (2018). Intellectual Property Rights – An Overview. Retrieved from http://www.bdu.ac.in/cells/ipr/docs/ipr-eng-ebook.pdf •World Intellectual Property Organization. (2004). WIPO Intellectual Property Handbook. (https://www.wipo.int/edocs/pubdocs/en/intproperty/489/wipo_pub	
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https://ww	v.springer.com/journal/10961
	v.ip.mpg.de/en/publications/journals/iic-international-review- al-property-and-competition-law.html
https://onli	nelibrary.wiley.com/journal/15406261
Journal Link:	

	M. TECH FIRST YEAR	
Course Code	AMTBT0251 L T P	Credit
Course Title	Bioinformatics Lab 0 0 4	2
Course objective		
1	To learn the various online databases	
2	To learn the online tools for analyzing various macromolecules of the cells	
3	To understand the phylogenetic analysis and related conclusions	
4	To learn the use of various tools for molecular analysis	
5	To understand the various methods for macromolecular sequencing	
Pre-requisites:		
	Students are expected to have knowledge of basic biology, cell biology and biochemistry	
Course Content	s / Syllabus	
1	To perform pair wise local and global sequence alignment for any two proteins and DNA sequences.	
2	To perform multiple sequence alignment for any five sequences and predicts the Phylogenetic relationship among them.	
3	Phylogenetic Analysis using PHYLIP - Rooted trees and Unrooted trees	
4	To predict secondary structure for any given protein sequence using Chou- Fasman, GOR and Neural network algorithms.	
5	To visualize tertiary structure of any given protein sequence using Rasmol/PyMol/PMV.	
6	To visualize the genomic map of Human genome and find out the size, number of genes and number of proteins encoded on Chr-Y.	
7	Homology Modelling using Modeller	
8	To find out the RMSD value from any two-protein structure alignment.	
9	Construction of Cladogram	
10	Different interactions using CYTOSCAPE	
11	Primary Structure Analysis of a Protein Using ProtParam	
12	Finding the Active Site Pockets of a given Protein Molecule	
Course outcome		
CO1	The students will learn the various online databases	
CO2	Students will learn the online tools for analysing various macromolecules of the cells	
CO3	They will understand the phylogenetic analysis and related conclusions	
CO4	The students will learn the use of various tools for molecular analysis	
CO5	The students will understand the various methods for macromolecular sequencing	

		M. TECH FIRST YEAR		
Course C	Code	AMTBT0252	L T P	Credit
Course T	itle	Entrepreneurship, IPR & Biosafety Lab	0 0 4	2
Course obj	iective			
1		To make students aware of the process of patent registration.		
2		To learn to design Biosafety lab.		
3		To develop the entrepreneur and marketing skills.		
Pre-requisi	ites:			
Students	are	expected to have knowledge of basic biology, cell biology and bio	chemistry	
<u> </u>				
Course Co	ntents	S / Syllabus		
1		Demonstration of Procedure for patent registration in India		
2		Writing a Patent Application		
3		Microbiological risk assessment		
4		Basic laboratories – Biosafety Levels 1 and 2 Basic laboratories Levels 1 and 2 Code of practice Laboratory design and facilities	– Biosafety	
5		Laboratory equipment Health and medical surveillance		
6		Biosafety practices and procedures		
7		Development of project proposals - SWOT analysis		
8		SWOT analysis of selected enterprise		
9		Practical on developing distribution channels; Pricing/Policies/ Promotion/Advertising through the use of social Media	Competition;	
10		Preparation of Balance Sheet		
Course out	tcome	1		
CO1		Students will be able to understand the process of establishing B labs.	iosafety	
CO2		Students will learn through demonstration the process Patent Re	gistration.	
CO3		They will develop the skills of marketing and entrepreneur.		

	M. TECH FIRST YEAR	
Course Code	AMTBT0211 L T P	Credit
Course Title	Genetic Engineering 3 0 0	3
Course objectiv	e:	
1	It is intended to impart basic undergraduate-level knowledge in the area of molecular biology and recombinant DNA technology.	
2	The student would be able to understand the working details of the cloning of a gene.	
3	They would also be able to assimilate recent research findings, advancement and development in the rDNA technology.	
4	The use of virtual lab and computational tools would enable them to perform in silico cloning of the selected DNA.	
5	To understand the DNA sequencing methods	
Pre-requisites:		
Students are expe	ected to have knowledge of basic biology, cell biology and biochemistry	
Course Content	s / Syllabus	
UNIT-I	Molecular Tools	8 Hr.
	Labelling of DNA: Nick translation, Random priming, Radioactive and non- radioactive probes, Hybridization techniques, Hybridization techniques; Chromatin Immunoprecipitation; DNA-Protein Interactions-Electromobility shift assay; DNaseIfootprinting; Methyl interference assay	
UNIT-II	Vectors	8 Hr.
	Plasmids; Bacteriophages; M13 mp vectors; PUC19 and Bluescript vectors, Phagemids; Lambda vectors, Insertion and Replacement vectors; Cosmids; Artificial chromosome vectors (YACs; BACs); Animal Virus derived vectors; Expression vectors; Inclusion bodies; Methodologies to reduce formation of inclusion bodies; Baculovirus and pichia vectors system, Plant based vectors, Ti and Ri as vectors, Yeast vectors, Shuttle vectors	
UNIT-III	Techniques in Genetic Engineering	8 Hr.
	Insertion of Foreign DNA into Host Cells; Transformation; Isolation of mRNA and total RNA; cDNA and genomic libraries and its construction; cDNA and genomic cloning; Expression cloning; Jumping and hopping libraries; Southwestern and Far-western cloning; Protein-protein interactive cloning and Yeast two hybrid system; Phage display; Principles in maximizing gene expression	
UNIT-IV	PCR and its applications	8 Hr.
	Primer design; Fidelity of thermostable enzymes; DNA polymerases; Concept of PCR, Types of PCR, Gene specific and degenerate primer design, linkers, adaptors, Fidelity of uDNA polymerase. Application of PCR. Chimeric protein engineering by PCR	

Course outco	Sequencing methods; Enzymatic DNA sequencing; Chemical sequencing of DNA; Automated DNA sequencing; RNA sequencing; Introduction of DNA into mammalian cells; Transfection techniques; Gene silencing techniques; siRNA technology; Micro RNA; Construction of siRNA vectors; Principle and application of gene silencing; Gene Therapy; Suicide gene therapy; Gene replacement; cDNA and intragenic arrays; Differential gene expression and protein array, Genome editing-CRISPR and other genome editing tools.	
Course outer	Understand the basic concept and procedure of gene cloning and the role of	
CO 1	enzymes and vectors used for genetic manipulation and genetic engineering	
CO 2	Acquired theoretical knowledge of vectors, their different types and applications in genetic engineering.	
CO 3	Getting detailed knowledge of construction of gene libraries and their screening methods.	
CO 4	Have knowledge of PCR technique, their different types and applications.	
CO 5	Understand the basic concept of genetic engineering techniques for selection of recombinants.	
Text books		
1	Winnacker, Ernst L. (1987), From genes to clones: introduction to gene technology [Gene und Klone] (in German), Horst Ibelgaufts (trans.), Weinheim, New York: VCH, ISBN 0-89573-614-4.	
2	Genetic Engineering by Dr Smita Rastogi & Dr Neelak Pathak, Oxford University Press	
3	Genetic Engineering, Principles& Practice by Sandhya Mitra, McGraw Hill Education.	
Reference Bo		
1	Principles of Gene Manipulation and Genomics, Primrose & Twyman.	
2	Molecular Biology of the Cell. 4th edition. Alberts B, Johnson A, Lewis J, et al. New York: Garland Science; 2002.	
3	Modern Genetic Analysis. Griffiths AJF, Gelbart WM, Miller JH, et al. New York: W. H. Freeman; 1999.	
Journal/Rese	earch Paper Link:	
	As suggested by concern subject faculty	

	M. TECH FIRST YEAR		
Course Code		L T P	Credit
Course Title	Applied Food Biotechnology	3 0 0	3
Course objectiv	e:		
1	To learn about the various microbiological examination of food born diseases	ls and food	
2	To learn about the development and production of novel products		
3	To understand GM foods and the legal issues associated with them	l.	
4	To learn about the industrial production of various food products		
5	To learn the methods of production of vitamins and enzymes.		
Pre-requisites:			
	Students are expected to have knowledge of basic biology, cell bio biochemistry	logy and	
Course Content	s / Syllabus		
Unit I	Food Biotechnology		8 Hr.
	Introduction & Applications; Methods for the microbiological ex of water and foods; Control of Microbiological quality and sa borne illnesses and diseases; Microbial cultures for food ferment maintenance, strain development	fety; Food	
Unit II	Biosensors in food technology		8 Hr.
	Starter cultures-types, designing and development, micro encapse packaging, scopes and challenge; Development and formulation products such as probiotic foods. Nutrigenomics-concept, significance and relevance. Biosensors and novel tools and their in food science & Technology	n of novel working,	
Unit III	GM foods		8 Hr.
	Introduction and controversies related to GMOs. Ethical issues GM foods; testing for GMOs; current guidelines for the producti and movement of GMOs; labelling and traceability; trade relate biosafety; risk assessment and risk management. Public perceptifoods. IPR. GMO Act–2004. New products and processes in vaccommodities including plant and animal products.	on, release ed aspects; ion of GM	
Unit IV	Industrial Food Biotechnology I		10 Hr.
	Industrial production of organic acids (vinegar, lactic acid), beverages (beer, wine, and distilled alcoholic beverages such a rum, vodka), glycerol; Propagation of baker's yeasts; Ferme products such as cheese, yoghurt, sweet curd, paneer, s Fermented pickles.	s whiskey, nted dairy	
Unit V	Industrial Food Biotechnology II		10 Hr.
	Industrial production of important primary and secondary metab as antibiotic, vitamins, biosurfantants, polysaccharides. Enzyme in food industry.Advantages and constraints of immobilized en microbial cells. Types of enzyme reactors. Aerobic and anaerobic of offluents from food processing industry.	application zymes and	
	of effluents from food processing industry		

To identify microorganism responsible for food spoilage.	
Demonstrate knowledge methods of packing, and the application of biosensors in food industries	
To understand the ethical issues lined with GM food production	
Demonstrate the industrial production of various food products	
To explain the industrial application of various enzymes	
Industrial Microbiology Prescott & Dunn, CBS Publishers	
Modern Food Microbiology by Jay JM, CBS Publishers	
Comprehensive Biotechnology by Murray &Mooyoung, Academic press	
Industrial Microbiology by Casida L.R., New Age International Pvt. Ltd.	
Food Microbiology; Frazier WC; 4th ed, Tata-McGrowhill Pub.	
ooks	
Microbiology by Pelczar, Chan, and Krieg, TMH	
Fermentation Biotechnology, Principles, Processed Products by Ward OP, Open	
University Press.	
Lee, B. H. Fundamentals of Food Biotechnology.VCH. 2006	
earch Paper Link:	
As suggested by concern subject faculty	
	biosensors in food industries To understand the ethical issues lined with GM food production Demonstrate the industrial production of various food products To explain the industrial application of various enzymes Industrial Microbiology Prescott & Dunn, CBS Publishers Modern Food Microbiology by Jay JM, CBS Publishers Comprehensive Biotechnology by Murray &Mooyoung, Academic press Industrial Microbiology by Casida L.R., New Age International Pvt. Ltd. Food Microbiology; Frazier WC; 4th ed, Tata-McGrowhill Pub. Microbiology by Pelczar, Chan, and Krieg, TMH Fermentation Biotechnology, Principles, Processed Products by Ward OP, Open University Press. Lee, B. H. Fundamentals of Food Biotechnology.VCH. 2006 arch Paper Link:

	M. TECH FIRST YEAR	
Course Code	AMTBT0213 L T P	Credit
Course Title	Molecular Modelling & Industrial Application3 0 0	3
Course objectiv	٥ .	
1	To learn about the basics of molecular modelling	
2	To understand the usage of computer simulation	
3	To understand the basic of drug development.	
4	To learn about the herbal drug and its trade scenario.	
5	To understand the method of vaccine production.	
Pre-requisites:	1	
	Students are expected to have knowledge of basic biology, cell biology and biochemistry	
Course Content	s / Syllabus	
Unit I	Molecular Modelling	8 Hr.
	Introduction; Useful Concepts in Molecular Modelling; The Molecular	0 111.
	Modelling Literature; Molecular Modelling software: BIOSUITE; Force Fields	
Unit II	Energy Minimisation and Computer Simulation	8 Hr.
	Minimisation and Related Methods for Exploring the Energy Surface. Non- Derivative method, 1st and 2nd order minimisation methods. Results of a Simulation and Estimating Errors. GROMACS and CNS. Molecular Dynamics & Monte Carlo Simulation.	
Unit III	Drugs	8 Hr.
	An introduction, Overview of drug discovery process, Trends in drug discovery process. Rationale of Drug Discovery: Medical needs, Target identification, Target validation, Receptors and assay development.	
Unit IV	Herbal Drugs	8 Hr.
	Definition, Trade scenario, Phytochemical standardization and fingerprinting, Marker compounds, Polyherbal formulations. Drug Development and Pre-Clinical Studies: Drug receptor interactions; enzyme inhibition and inactivation, In-vitro and in-vivo pharmacodynamic models, Therapeutic index, Pharmacokinetics - Microbial and animal models, In- vitro and insilico toxicological models, Drug formulations.	
Unit V	Applications of microbes for designing vaccines	8 Hr.
	Applications of microbes for designing vaccines: case study.	
Course outcome	2	
1	Students will learn about the basics of molecular modelling	
2	Students will understand the usage of computer simulation	
3	Students will understand the basic of drug development.	
4	Students will learn about the herbal drug and its trade scenario.	
5	Students will be able to understand the method of vaccine production.	
Text books		

1	A.R.Leach, Molecular Modelling Principles and Application, Longman, 2001.		
2	J.M.Haile, Molecular Dynamics Simulation Elementary Methods, John Wiley and Sons, 1997.		
3	Satya Prakash Gupta, QSAR and Molecular Modelling, Springer - Anamaya Publishers, 2008.		
4	Patwardhan B, Drug Discovery and Development-Traditional Medicine and Ethnopharmacology, New India Publishing (2007)		
5	Larsen PK, Leljifore T and Medsan U, Text Book of Drug Design and Discovery, CRC Press (2009)		
Reference Book	XS		
1	Hillisch A and Hilgenfeld R, Modern Methods of Drug Discovery, Birkhauser (2003).		
Journal/Research Paper Link:			
	As suggested by concern subject faculty		

M. TECH FIRST YEAR				
Course Code	AMTBT0214 L T P	Credit		
Course Title	Bioreactor Analysis and Design3 0 0	3		
Course objectiv				
1	To learn about the designing of bioreactor systems			
2	To learn about the control involved in bioreactor system			
3	To learn about the various types of bioreactor processes			
4	To understand the reactor dynamics			
5	To learn the design aspect and safety issues.			
Pre-requisites:				
	Students are expected to have knowledge of basic biology, cell biology and biochemistry			
Course Content	s / Syllabus			
UNIT I	Material balance and design	8 Hr.		
	Introduction; General design information; Material and energy balance calculations; Process Flow sheeting,Selection of bioprocess equipment (upstream and downstream); Specifications of bioprocess equipment; Mechanical design of reactors, heat transfer and mass transfer equipment; Design considerations for maintaining sterility of process streams and process equipment; Piping and instrumentation; Materials of construction for bioprocess plants.			
UNIT II	Control of bioreactor	8 Hr.		
	Basic aspects of bioreactor designing, Physical, chemical and biological sensors and control, Advanced control strategies viz. PID controllers, Fuzzy logic based controllers and Artificial Neural Network (ANN) based controllers, Basic concepts of computer modelling and optimization in bioprocess applications.			
UNIT -III	Ideal Bioreactor and its working	8 Hr.		
	Ideal bioreactors: Batch reactors, Fed-batch reactors, enzyme-catalyzed reaction in CSTRs, CSTR reactors with recycle and wall cell growth, the ideal plug-flow tubular reactor, Reactors with nonideal mixing: Mixing times in agitated tanks, residence time distribution, models for nonideal reactors, Mixing-bioreaction interactions.			
UNIT -IV	Types of Bioreactors	8 Hr.		
	Reactor dynamics and stability, Multiphase bioreactors: conversion of heterogeneous substrates, packed-bed reactors, bubble column bioreactors, fluidized bed bioreactors, trickle-bed reactors, airlift reactor, Immobilized Enzyme reactors, Photo bioreactors, Hollow fibre membrane bioreactors. Scale up and scale down issues: Effect of scale on oxygenation, mixing, sterilization, pH, temperature, inoculum development, nutrient availability and supply; Bioreactor scale-up based on constant power consumption per volume, mixing time, impeller tip speed (shear), mass transfer coefficients.			

UNIT V	Downstream Processing	8 Hr.
	Facility design aspects; Utility supply aspects; Equipment cleaning aspects;	
	Culture cell banks; cGMP guidelines; Validation; Safety. Process	
	economics; Case studies, Scale up of downstream processes: Adsorption (LUB method); Chromatography (constant resolution etc.); Filtration	
	(constant resistance etc.); Centrifugation (equivalent times etc.); Extractors	
	(geometry based rules).	
Course outco		
CO1	After completing the course students will able to design the bioreactor system	
CO2	After completing the course students will able to illustrate the control involved in bioreactor system	
CO3	After completing the course students will able to identify the various types of bioreactor processes	
CO 4	After completing the course students will able to analyse the reactor dynamics	
CO5	After completing the course students will able to evaluate the design aspect and safety issues associated with reactor system.	
Text books		
1	Moser, Anton, Bioprocess Technology: Kinetics and Reactors, Springer Verlag, 1988.	
2	Bailey J.E. &Ollis, D.F. Biochemical Engineering Fundamentals, 2nd ed., McGraw Hill, 1986	
3	Lee, James M. Biochemical Engineering, PHI, USA.	
4	Atkinson, Handbook of Bioreactors, Blanch, H.W. Clark, D.S. Biochemical Engineering, Marcel Decker, 1999	
5	Biochemical Engineering fundamentals" 2nd edJ E Bailey and D F Ollis, McGraw-Hill (1986) Chapters 8,9&10.	
6	Biochemical Engineering" -S Aiba, A E Humphrey and N Millis , 1978, University of Tokyo Press.	
7	Biotechnology" Vols. 3 & 4 Eds., S Rehm and G Reed. VCH (1991).	
Reference B	ooks	
1	Biochemical Engineering and Biotechnology Handbook" 2nd Ed., Atkinson & F.Mavituna, Stockton Press (1991).	
2	Biorector Design & Product Yield", BIOTOL series, Butterworth - Heinemann (1992).	
3	Principles of fermentation technology" - F Stanbury and A Whitaker, Pergamon press (1984)	
4	Unit operations of Chemical Engineering" 5th ed. by W L McCabe, J C Smith and P. Harriot Mc Graw-Hill (1993).	
5	Bioprocess Engineering Principles" by Pauline M.Doran, Academic Press.	
6	Feedback and Control systems- Schaum's outline series, McGraw-Hill Book Comp., 1967	
7	Unit Operations of Chemical Engineering- Mc Caba Smith, Harriott, Mc Graw – Hill Chemical Engg. Series., V Ed., 1985.	
Journal/Reso	earch Paper Link:	
	As suggested by concern subject faculty	

	M. TECH FIRST YEAR	
Course Code	AMTBT0215 L T P	Credit
Course Title	Enzyme Technology & Industrial Application 3 0 0	3
Course objectiv	e:	
1	To learn about the kinetics involved in enzymatic reactions.	
2	To learn about the various biochemical processes involved in the microbial growth	
3	To learn about the various processes in bioreactor	
4	To understand the various separation methods involved in bioprocess	
5	To analyze the different bioprocess steps in industrial production.	
Pre-requisites:		
	Students are expected to have knowledge of basic biology, cell biology and biochemistry	
Course Content	s / Syllabus	
UNIT I-	ENZYME TECHNOLOGY	8 Hr.
	Introductions: Enzymes- Michaelis-Menten kinetics. Kinetics and StatisticsInhibition- Effect of pH and temperature- Enzymology- Immobilized enzymes: Methods, Mass transfer considerations and Industria enzymes.	-
UNIT II-	METABOLISM, STOICHIOMETRY AND MICROBIAL GROWTH KINETICS	
	Introduction to metabolism- Nutrient transport- Glycolysis - TCA cycle and other pathways - Control of metabolism. Factors affecting microbial growth – Stoichiometry- mass balances and energy balances. Growth kinetics Measurement of growth.	1
	BIOREACTORS, STERILIZATION, SENSORS AND	0.11
UNIT III-	INSTRUMENTATION	8 Hr.
	Introduction to bioreactors - Batch and Fed-batch bioreactors, Continuous bioreactors, Immobilized cells, Bioreactor operation, Sterilization, Aeration Sensors. Instrumentation, Culture - specific design aspects plant/mammalian cell culture reactors.	,
UNIT IV-	PRIMARY & SECONDARY SEPARATION PROCESS	8 Hr.
	Biomass removal - Biomass disruption – Membrane based techniques Extraction -solvent, aqueous two phases, super critical, and Adsorption Chromatography, Precipitation (Ammonium Sulfate, solvent) Electrophoresis (capillary), Crystallization, Drying and Freeze drying.	
UNIT V-	INDUSTRIAL APPLICATION	8 Hr.

	White Biotechnology: Few industrial process using enzymes for productionof drugs and fine chemicals, Enzyme based biosensors, Enzyme in organiccatalysis,Analyticalapplications, Applicationsinfood	
	industry,Pharmaceuticals,Biochemical applications: Role of soluble and immobilized enzymes in the synthesis and production of amino acids and chiral compounds; use of enzymes as detergents.Molecular Imprinting; Enzyme engineering: <i>In vitro</i> approaches to improve functional efficiency; Recombinant enzymes, Case study.	
Course out	come	
CO1	Describe the fundamentals of enzyme properties, nomenclatures, characteristics and mechanisms & plot graphs based on kinetics data.	
CO2	Demonstrate metabolism, stoichiometry and microbial growth kinetics.	
CO3	Perform bioreactor operations as applicable in bioprocess industries.	
CO4	Discuss various separation and purification process of fermentation products.	
CO5	Predict the current and future trends of applying enzyme technology for the commercialization purpose of biotechnological products.	
Text books		
1	Michael Shuler and FikretKargi. "Bioprocess Engineering: Basic Concepts", 2nd Edition, Prentice Hall, and Englewood Cliffs, NJ, 2002.	
2	Pauline Doran. "Bioprocess engineering principles", Academic Press, 1995.	
3	Colin Ratledge, Bjorn Kristiansen, "Basic Biotechnology", 2nd Edition, Cambridge University Press, 2001.	
Reference I	Books	
1	Roger Harrison et al., "Bioseparation Science and Engineering", Oxford University Press, 2003.	
2	Harrison R.G. Todd P., Rudge S.R. "Bioseparation Science and Engineering", Oxford Press 2003.	
Journal/Re	search Paper Link:	
	As suggested by concern subject faculty	

Course Code Course Title Course objective	AMTBT0216L T PApplied Bioenergy3 0 0	Credit
	Applied Bioenergy 3 0 0	
Course objective		3
	•	
1	To understand the basics of bioenergy	
2	To learn the principals of biofuel production	
3	To learn about the current application of bioenergy	
4	To understand the impact of energy on economy	
5	To understand production of biofuels in real life.	
Pre-requisites:		
	Students are expected to have knowledge of basic biology, cell biology and biochemistry	
Course Contents	s / Syllabus	
Unit I	Introduction to applied bioenergy	8 Hr.
	Introduction to applied bioenergy, Types of bioenergy, Energy scenario-role of energy in economic development and social transformation, Commercial and non-commercial forms of energy, Present and future global projections of energy consumptions.	
Unit II	Biomass and Energy Conservation	8 Hr.
	Principles of biomass energy conversion processes, biological, chemical and thermo-chemical technologies for biomass conversion and their utilization covering: Biogas, Produces gas, Alcohol and Biodiesel, Second generation biofuel from high efficiency algal-derived biocrude, Biobased fats (Lipids) and oils from biomass for energy production, Biorefinery systems: An Overview. Microbial fuel cell and their application	
Unit III	Bioenergy	8 Hr.
	Current bio-energy applications and conversion technologies, Advantages of applied bioenergy over other sources of energy, Advances in bio-energy research: An overview of technological developments, bioenergy value chain, Databases of bioenergy related enzymes, Sustainable farming of bioenergy crops.	
Unit IV	Impact of Energy on Economy and Environment	8 Hr.
	Impact of Energy on Economy, Development and Environment, Energy for Sustainable Development, Energy and Environmental policies, Need for use of new and renewable energy sources, Energy Policy Issues: Fossil Fuels, Renewable Energy, Power sector reforms, restructuring of energy supply sector, energy strategy for future, Status of Nuclear and Renewable Energy: Present Status and future promise.	
Unit V	Case study	8 Hr.
	Case study 1: Biodiesel from Jatropha plant as transport fuel, A case study of UP State (India) 2. Generation of Bio-fuel by Using Waterweeds: A Case Study in Solapur City	

CO1	Demonstrate different types of bioenergy.			
CO2	Demonstrate the production of various types of biofuel using different substrates.			
CO3	To explain the advantages of applied bioenergy over other sources of energy and advances in bio-energy research.			
CO4	To describe the Impact of Energy on Economy.			
CO5	To describe the application of of biofuel in real life.			
Text books				
1	Anthony San Pietro (1980); Biochemical and Photosynthetic aspects of Energy Production, Academic Press, New York.			
2	Berman, ER Geothermal Energy, Noyes Data Corporation, New Jersey			
3	Parker, Colin & Roberts, (1985); Energy from Waste- An Evaluation of Conversion Technologies, Elsevier Applied Science London			
Reference	Books			
1	Ralph E.H. Simsed. (2004); Bioenergy options for cleaner environment by World Renewable Energy Network.			
2	Ravindranath N.H. and Hall D.O. (1995); Biomass, Energy and Environment, A developing country perspective from India by, Oxford University Press,			
3	Brown Robert C. (2003); Biorenewable Resources: Engineering New Products from Agriculture, Iowa State University Press ,USA			
4	Boyle Godfrey ed. (1996): Renewable Energy: Power for a sustainable future, Oxford, OUP			
Journal/Re	esearch Paper Link:			
	As suggested by concern subject faculty			

		M. TECH FIRST YEAR		
Course	Code	AMTBT0217	L T P	Credit
Course	Title	Cell & Tissue Culture Techniques	300	3
Course o	bjectiv	e:		
1	3	To learn the basics of animal cell culturing technique.		
2		To understand the various methods and advancements of culture tee	chniques.	
3		To analyse the applications of animal cell culturing.		
4		To learn the basics of plant cell and tissue culture technique.		
5		To understand the various methods and advancements in plant cell culture.	and tissue	
Pre-requ	isites:			
Course (Content	Students are expected to have knowledge of basic cell and molecula s / Syllabus	ar biology.	
Unit 1		z Tissue Culture Technology Basics		8 hr
	Physic Tempo	cell culture techniques, Types of cell culture media; Ingredients ochemical properties; CO_2 and bicarbonates; Buffering; Oxygen; Oterature; Surface tension and foaming; Balance salt solutions; An supplements;	smolarity;	
Unit 2	Metho	ods of Cell & Tissue Culture		8 hr
	fibrob Trypsi culture	ent tissue culture techniques; Types of primary culture; Chicken last culture; Chicken liver and kidney culture; Secondary nization; Cell separation; Continuous cell lines; Suspension cultu e etc.; Behaviour of cells in culture conditions: division, growt olism of estimation of cell number; Development of cell lines	culture; re; Organ	
Unit 3	Appli	cations of Cell and Tissue Culture Technique		8 hr
	Cell cloning and selection; Transfection and transformation of cells; Commercial scale production of animal cells, stem cells and their application; Application of animal cell culture for <i>in vitro</i> testing of drugs; Testing of toxicity of environmental pollutants in cell culture; Application of cell culture technology in production of human and animal viral vaccines and pharmaceutical proteins, Green Meat, Organ Printing			
Unit 4	Plant	Cell & Tissue Culture Basics		10 hr
	embry improv diseas and c	mentals of plant tissue culture, plant regeneration: organogenesis ogenesis; somaclonal variation, its genetic basis and application vement. Cell/callus line selection for resistance to herbicide, s es.: Isolation, culture and plant regeneration, protoplast fusion, iden haracterization of somatic hybrids., Field techniques for propa erated plants.	n in crop stress and ntification	
Unit 5	Techn	iques of Plant Cell & Tissue Culture		10 hr
	B5, S param	nt selection, sterilization and inoculation; Various media preparate 5H PC L2; Callus and cell suspension culture; Induction an eters; Chromosomal variability in callus culture. Plant regenera o, meristem and callus culture. Androgenesis: Anther and pollen cul-	d growth tion from	

Course	outcome:		
CO1	After completion of the course, students will learn the basics of animal cell culturing.		
CO2	They will understand about the various methods and protocols of cell culturing.		
CO3	They will analyse the different types of applications of animal cell culturing.		
CO4	Students will learn the basics of plant tissue culture.		
CO5	Students will be able to understand the different methods of plant tissue culture and their applications.		
Text be	ooks		
1	B. Hafez and E.S.E Hafez, Reproduction in farm animals, 7th Edition, Wiley Blackwell, 2000		
2	G.E. Seidel, Jr. and S.M. Seidel, Training manual for embryo transfer in cattle (FAO Animal Production and Health Paper-77), 1st Edition, W.D. Hoard and sons FAO, 1991		
3	I. Gordon, Laboratory production of cattle embryos, 2nd edition, CAB International, 2003.		
Refere	nce Books		
1	Louis-Marie Houdebine, Transgenic Animals: Generation and Use 5th Edition, CRC Press, 1997.		
2	Plant Tissue Culture: Theory and Practice, a Revised Edition by S.S. Bhojwani and M.K. Razda		
3	Plants from Test Tubes: An Introduction to Micropropagation by LydianeKyte		
Journa	I/Research Paper Link:		
	As suggested by concern subject faculty		

M. TECH FIRST YEAR				
Course Code	AMTBT0218 L T 1	P Credi		
Course Title	Diagnostic Techniques in Biotechnology 3 0 () 3		
Course objective	2:			
1	To learn the basics of diagnostic techniques.			
2	To understand the different enzymes and related test methods.			
3	To learn the methods of immunodiagnostics.			
4	To understand the product development related to diagnostics.			
5	To learn the methods of DNA based diagnostics.			
Pre-requisites:				
*	Students are expected to have knowledge of basic biology, cell biology at biochemistry	nd		
Course Contents	s / Syllabus			
Unit I	Analytical Methods	8 hr		
	Volumetric analysis, Balancing & Weighing, Concept of solute & solve Units of measurement. Specimen Collection & Processing: Specim collection (Blood, urine, spinal fluid, saliva synovial fluid, Amniotic fluid Preservation, transportation	nen		
Unit II	Clinical Enzymology	8 hr		
	Principle of diagnostic enzymology, Digestive enzyme, Miscellaned enzyme. General Function Tests: Liver function test, Cardiac Function Te Renal Function Test, Thyroid Function test, Reproductive endocr function test	est,		
Unit III	Immunodiagnostics	8 hr		
	Introduction, Antigen-Antibody Reactions, Conjugation Techniqu Antibody Production, Enzymes and Signal Amplification System Separation and Solid-Phase Systems, Studies related to bacterial, viral a parasitic infections.	ms,		
Unit IV	Product Development	10 hr		
	Immunoassay Classification and Commercial Technologies, Ass Development, Evaluation, and Validation, Reagent Formulations and Sh Life Evaluation, Data Analysis, Documentation, Registration, a Diagnostics Start-ups.			
Unit V	DNA based diagnostics	10 hr		
	PCRRT-PCR, qPCR, Hot start PCR, Nested PCR), RFLP, SSC Microarrays, FISH, In-situ hybridization, Studies related to bacterial, vi and parasitic infections, Cell based diagnostics: Antibody markers, O Markers, FACS, HLA typing, Bioassays, Viral DNA detection using Rakits and PCR	iral CD		
Course outcome				
CO1	The students will learn the basics of diagnostic techniques.			
CO2	The students will understand the different enzymes and related test method	ods.		
CO3	The students will learn the methods of immunodiagnostics.			

CO4	The students will understand the product development related to diagnostics.	
CO5	The students will learn the methods of DNA based diagnostics.	
Text books		
1	Tietz Textbook of Clinical Chemistry, Carl A. Burtis, Edward R. Ashwood, Harcourt Brace & Company AisaPvt. Ltd.	
2	Commercial Biosensors: Graham Ramsay, John Wiley & Son, INC. (1998).	
3	Essentials of Diagnostic Microbiology, Lisa Anne Shimeld.	
Reference Books	S S	
1	Diagnostic Microbiology, Balley& Scott's.	
2	Tietz Text book of Clinical Biochemistry, Burtis& Ashwood. 6. The Science of Laboratory Diagnosis, Crocker Burnett.	
Journal/Researc	h Paper Link:	
	As suggested by concern subject faculty	

		M. TECH FIRST YEAR	-	
Course	Code	АМТВТ0219	L T P	Credit
Course Title 3-D Printin		3-D Printing Technology	300	3
Course of	ojective	:		
		know the fundamentals of RP Systems & its evolution and the Procession of RP Systems with 3D modelling & Mesh	cocess in RP	K1,K2
2	Able to	know the RP Systems, Process, Materials & Classifications		K3, K4
3	format,	b know and working with Mesh File & their formats like STL for OBJ formats. Conversion to Mesh files, their properties, operation ions & defects		K3, K4
4	Able to	know the applications of RP Systems in various Fields		K3, K4
Pre-requi	sites:			
Basic und	erstandi	ng of Information Technology.		
Course C	ontents	s / Syllabus		
UNIT-I	Intro	duction:		4 hours
	differ	rical Developments, Fundamentals of RP Systems and its Class rent basis, Rapid Prototyping Process Chains, 3D Modelling ration, Data Conversion and Transmission.		
UNIT-II	RP S	ystems:		12 hours
	Liquid Polymer Based Rapid Prototyping systems: SLA, Material Jetting, Solid Input Materials Based Rapid Prototyping Systems: Laminated Object Manufacturing (LOM) and Fused Deposition Modelling Systems, Power Based Rapid Prototyping Systems: Selective Laser Sintering, Multi-Jet Fusion, Binder Jetting Systems.			
UNIT-III		Database & Design Optimization:		8 hours
		d Prototyping Data Formats, STL Format, STL file problems, STI M, Topology Optimization, Gcode for RP Systems	file repair,	
UNIT-IV	RP A	pplications:		8 hours
	Development of dies for Moulding, RP Applications in developing prototypes of products, application in medical fields, Development of bone replacements and tissues, etc., RP materials and their biological acceptability.			
Course ou	itcome	After completion of this course students will be able to		
CO 1	Under	stand the fundamentals of RP Technologies and process involvement	ent in them	K1,K2
CO 2		Understand the methodology to manufacture the products using RP technologies and study their applications, advantages and case studies		K3, K4
CO 3	Understand the Design aspects and their respective challenges along with the resolution for them		K3, K4, K5	
CO 4	Understand the various applications of various RP Systems with case studies & Materials		K3,K4	
Text book	KS			
1	Rapid Prototyping: Principles an Applications: Chee Kai Chua, Kah Fai Leong, Chu Sing Lim			
2	Additive Manufacturing Technologies: 3D Printing, Rapid Prototyping, and Direct Digital Manufacturing: Brent Stucker, David W. Rosen, Ian Gibson			

Reference Books		
1	Rapid Manufacturing: The Technologies and Applications of Rapid Prototyping and Rapid Tooling: Pham, Duc, Dimov, S.S.	
2	Rapid Prototyping and Manufacturing: Fundamentals of Stereo Lithography: P. Jacobs	
3	Rapid System Prototyping with FPGAs: Accelerating the Design Process: R.C. Cofer, Benjamin F. Harding	
4	Rapid Prototyping of Digital Systems: Hamblen, James O., Hall, Tyson S., Furman, Michael D.	